

**Polycaspase Live!, Green and Red-Catalog#s: KF17303 and KF17304  
Protocol**

1. Expose your animals to your experimental condition, and create positive and negative controls if possible.
2. Reconstitute the reagent with 50uL DMSO to form the stock concentrate (which can be frozen for future use).
3. Dilute the injection buffer 1:10 with diH<sub>2</sub>O and sterilize by filtration. Add 200uL 1X injection buffer to the reagent.
4. Inject ~40uL of diluted reagent into each mouse.
5. Let circulate 30-45 minutes.
6. Examine tissues under a fluorescent microscope, or sacrifice and excise cells.
7. If desired, label cells with an additional stain, like red fluorescent vital stain or labeled antibody.
8. If desired, fix, embed, or freeze cells.
9. Analyze cells with a fluorescent microscope, plate reader or flow cytometer.

Note: This protocol may be adjusted depending on your animal and target tissues. For example, to increase the dose of Polycaspase Live! given to each animal, change steps 4 and 5. In step 4, add 550uL injection buffer to the reagent (creating 600 uL of injection solution), then in step 5, inject 100uL into each animal.

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**12/06v1**

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