

## Polyclonal Antibody to p38 MAP Kinase pThr180/pTyr182 - Aff - Purified

<b>Catalog No.:</b>	BP7136
<b>Quantity:</b>	0.1 ml
<b>Background:</b>	p38 is a 38 kDa Stress Activated Protein Kinase/Map Kinase (SAPK/MAPK) represented by three isoforms, termed $\alpha$ , $\beta$ , and $\gamma$ . p38 is dually phosphorylated and therefore fully activated by MEK3 and MEK6 on threonine 180 and tyrosine 182 within the activation loop. The JNK and p38 pathways function to modulate cell-cycle, apoptotic and transcriptional responses to stress.
<b>Host:</b>	Rabbit
<b>Immunogen:</b>	Chemically synthesized phosphopeptide derived from the region of human p38 that contains threonine 180 and tyrosine 182. <b>Remarks:</b> This region is conserved among many species including mouse, rat, dog, monkey and carp.
<b>Format:</b>	<b>State:</b> Liquid Ig fraction <b>Purification:</b> Sequential epitope-specific chromatography. The antibody has been negatively preadsorbed using a non-phosphopeptide corresponding to the site of phosphorylation to remove antibody that is reactive with non-phosphorylated p38. The final product is generated by affinity chromatography using a p38-derived peptide that is phosphorylated at threonine 180 and tyrosine 182. <b>Buffer System:</b> Dulbecco's phosphate buffered saline (without Mg <sup>2+</sup> and Ca <sup>2+</sup> ), pH 7.3 (+/- 0.1), 50% glycerol, with 1.0 mg/mL BSA (IgG, protease free) as a carrier, containing 0.05 % sodium azide as preservative
<b>Applications:</b>	Western blot (1:1000). Positive Controls Used: HEK 293 cells +/- UV irradiation treatment and PC12 cells +/- Sorbitol. Other applications not tested. Optimal dilutions are dependent on conditions and should be determined by the user.
<b>Specificity:</b>	This antibody detects p38 $\alpha$ , $\beta$ and $\gamma$ . <b>Species:</b> Human, Mouse, Rat, Dog, Monkey, Carp. Other species not tested.
<b>Storage:</b>	Store the antibody at -20 °C. Can be shipped at 2 - 8 °C. Avoid repeated freezing and thawing. Centrifuge vial before opening. Shelf life: One year from despatch.
<b>General Readings:</b>	Karasewski, L. and A. Ferreira (2003) MAPK signal transduction pathway mediates agrin effects on neurite elongation in cultured hippocampal neurons. J. Neurobiol. 55(1):14-24.

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Nanki, T., et al. (2001) Chemokines regulate IL-6 and IL-8 production by fibroblast-like synoviocytes from patients with rheumatoid arthritis. *J. Immunol.* 167(9):5381-5385.

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Marinissen, M.J., et al. (2001) Regulation of gene expression by the small GTPase Rho through the ERK6 (p38 gamma) MAP kinase pathway. *Genes Dev.* 15(5):535-553.

Nelson, J.M. and D.W. Fry (2001) Akt, MAPK (Erk1/2), and p38 act in concert to promote apoptosis in response to ErbB receptor family inhibition. *J. Biol. Chem.* 276(18):14842-14847.

Zhang, H., et al. (2001) Stress-induced inhibition of ERK1 and ERK2 by direct interaction with p38 MAP kinase. *J. Biol. Chem.* 276(10):6905-6908.

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**Protocols:****Western Blotting Procedure**

1. Lyse approximately 10e7 cells in 0.5 mL of ice cold Cell Lysis Buffer (formulation provided below). This buffer, a modified RIPA buffer, is suitable for recovery of most proteins, including membrane receptors, cytoskeletal-associated proteins, and soluble proteins. Other cell lysis buffer formulations, such as Laemmli sample buffer and Triton-X 100 buffer, are also compatible with this procedure. Additional optimization of the cell stimulation protocol and cell lysis procedure may be required for each specific application.
2. Remove the cellular debris by centrifuging the lysates at 14,000 x g for 10 minutes. Alternatively, lysates may be ultracentrifuged at 100,000 x g for 30 minutes for greater clarification.
3. Carefully decant the clarified cell lysates into clean tubes and determine the protein concentration using a suitable method, such as the Bradford assay. Polypropylene tubes are recommended for storing cell lysates.
4. React an aliquot of the lysate with an equal volume of 2x Laemmli Sample Buffer (125 mM Tris, pH 6.8, 10% glycerol, 10% SDS, 0.006% bromophenol blue, and 130 mM dithiothreitol [DTT]) and boil the mixture for 90 seconds at 100°C.
5. Load 10-30 µg of the cell lysate into the wells of an appropriate single percentage or gradient minigel and resolve the proteins by SDS-PAGE.
6. In preparation for the Western transfer, cut a piece of PVDF membrane slightly larger than the gel. Soak the membrane in methanol for 1 minute, then rinse with ddH<sub>2</sub>O for 5 minutes. Alternatively, nitrocellulose may be used.
7. Soak the membrane, 2 pieces of Whatman paper, and Western apparatus sponges in transfer buffer (formulation provided below) for 2 minutes.
8. Assemble the gel and membrane into the sandwich apparatus.
9. Transfer the proteins at 140 mA for 60-90 minutes at room temperature.
10. Following the transfer, rinse the membrane with Tris buffered saline for 2 minutes.
11. Block the membrane with blocking buffer (formulation provided below) for one hour at room temperature or overnight at 4°C.
12. Incubate the blocked blot with primary antibody at a 1:1000 dilution in Tris buffered saline supplemented with 3% BSA and 0.1% Tween 20 overnight at 4 °C or for one hour at room temperature.
13. Wash the blot with several changes of Tris buffered saline supplemented with 0.1% Tween 20.
14. Detect the antibody band using an appropriate secondary antibody, such as goat F(ab)<sub>2</sub> anti-rabbit IgG alkaline phosphatase conjugate or goat F(ab)<sub>2</sub> anti-rabbit IgG horseradish

peroxidase conjugate in conjunction with your chemiluminescence reagents and instrumentation.

**Cell Lysis Buffer Formulation:**

10 mM Tris, pH 7.4  
100 mM NaCl  
1 mM EDTA  
1 mM EGTA  
1 mM NaF  
20 mM Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>  
2 mM Na<sub>3</sub>VO<sub>4</sub>  
0.1% SDS  
0.5% sodium deoxycholate  
1% Triton-X 100  
10% glycerol  
1 mM PMSF (made from a 0.3 M stock in DMSO)  
or 1 mM AEBSF (water soluble version of PMSF)  
60 µg/mL aprotinin  
10 µg/mL leupeptin  
1 µg/mL pepstatin  
(alternatively, protease inhibitor cocktail such as Sigma Cat. # P2714 may be used)

**Transfer Buffer Formulation:**

2.4 gm Tris base  
14.2 gm glycine  
200 mL methanol  
Q.S. to 1 liter, then add 1 mL 10% SDS.  
Cool to 4°C prior to use.

**Tris Buffered Saline Formulation:**

20 mM Tris-HCl, pH 7.4  
0.9% NaCl

**Blocking Buffer Formulation:**

100 mL Tris buffered saline  
5 gm BSA  
0.1 mL Tween 20

**Peptide Competition Experiment**

The specificity of a Phosphorylation Site Specific Antibody (PSSA) in each experimental system can be confirmed through peptide competition. In this technique, aliquots of antibody are pre-incubated with peptide containing the sequence of the phosphopeptide immunogen used to raise the PSSA and the corresponding non-phosphopeptide. Following preincubation with the peptide, each antibody preparation is then used as a probe in antibody-based detection methods, such as Western blotting, immunocytochemistry, flow cytometry, or ELISA. With a PSSA specific for the phosphorylated target protein, pre-incubation with an excess of peptide containing the sequence of the phosphopeptide immunogen will block all antigen binding sites, while pre-incubation with the corresponding non-phosphopeptide will not affect the antibody.

In performing the Peptide Competition Experiment, it is important to note that the optimal

dilutions of both antibody and peptide should be determined empirically for each specific application. The optimal dilution of antibody in these procedures is below saturating, as determined by previous experiments in your system.

The optimal dilution of peptide used in these procedures will depend on the overall affinity or avidity of the antibody, as well as the quantity of the target antigen. A 50-150 fold molar excess of peptide to antibody is found to be effective for most peptide competition experiments.

In the example presented below, the PSSA is used as a dilution of 1:1000 and the peptides are used at a concentration of 333 nM. The total volume of the phosphopeptide and nonphosphopeptide pre-incubated antibody preparations is 2 mL, sufficient for probing Western blot strips, as well as for use in other antibody-based detection methods. Under these conditions, the molar excess of peptide to antibody is  $> / = 50$ .

#### Procedure:

1. Prepare three identical test samples, such as identical PVDF or nitrocellulose strips to which the protein of interest has been transferred. The test samples should be blocked using a blocking buffer, such as Tris buffered saline supplemented with 0.1% Tween 20, and either 5% BSA or 5% non-fat dried milk.
  2. Prepare 6.5 mL of working antibody stock solution (1:1000 in this example) by adding 6.5  $\mu$ L of antibody stock solution to 6.5 mL of buffer containing blocking protein, such as TBS supplemented with 0.1% Tween 20, and either 3% BSA or 3% non-fat dried milk.
  3. Apportion the unused PSSA into working aliquots and store at  $-20^{\circ}\text{C}$  for future use (the stock PSSA contains 50% glycerol and will not freeze at this temperature).
  4. Allow the lyophilized control peptides to reach room temperature, ideally under desiccation.
  5. Reconstitute each of the control peptides to a concentration of 66.7  $\mu\text{M}$  with nanopure water. (i.e. for a peptide with a molecular mass of 1500, reconstitution with 1 mL water yields a solution with a concentration of 66.7  $\mu\text{M}$ ).
  6. Apportion the unused reconstituted peptide solutions into working aliquots and store at  $-20^{\circ}\text{C}$  for future use.
  7. Label 3 test tubes as follows:
    - tube 1: water only no peptide control
    - tube 2: phosphopeptide
    - tube 3: non-phosphopeptide
  8. Into each tube, pipette the following components
    - tube 1: 2 mL diluted PSSA solution plus 10  $\mu$ L nanopure water
    - tube 2: 2 mL diluted PSSA solution plus 10  $\mu$ L phosphopeptide
    - tube 3: 2 mL diluted PSSA solution plus 10  $\mu$ L non-phosphopeptide
  9. Incubate the three tubes for 30 minutes at room temperature with gentle rocking. During this incubation, the peptides have the chance to bind to the combining site of the antibody.
  10. At the end of the incubation step, transfer the contents of each of the three tubes to clean reaction vessels containing one of the three identical test samples.
- For Western blotting strips:  
Incubate the strips with the pre-incubated antibody preparations for 1 hour at room temperature or overnight at  $4^{\circ}\text{C}$ .  
Wash each strip four times, five minutes each, to remove unbound antibody. Transfer each strip to a new solution containing a labeled secondary antibody [e.g., goat F(ab)<sub>2</sub> anti-rabbit IgG alkaline phosphatase conjugate or goat F(ab)<sub>2</sub> anti-rabbit IgG horseradish peroxidase conjugate].  
Remove unbound secondary antibody by thorough washing, and develop the signal using your chemiluminescent reagents and instrumentation.  
The signal obtained with antibody incubated with the "Water Only, No Peptide Control"

(Tube 1), represents the maximum signal in the assay. This signal should be eliminated by preincubation with the "Phosphopeptide" (Tube 2), while pre-incubation with the "Non-Phosphopeptide" (Tube 3) should not impact the signal. If the "Phosphopeptide" only partially eliminates the signal, repeat the procedure using twice the volume of water or peptide solutions listed in Step 8. If partial competition is seen following pre-incubation with the "Non-Phosphopeptide", repeat the procedure using half the volumes of water or peptide solutions listed in Step 8.

**Pictures:**

**Peptide Competition** Extracts prepared from HEK 293 cells treated with UV irradiation were resolved on a 10% polyacrylamide gel and transferred to PVDF. Membranes were blocked with a 5% BSA-TBST buffer overnight at 4°C, then were incubated with p38 [pTpY180/182] antibody for two hours at room temperature in a 3% BSA-TBST buffer, following its prior incubation with: the peptide immunogen (1), a generic phosphothreonine-containing peptide (2), a generic phosphotyrosine-containing peptide (3), the non-phosphorylated peptide corresponding to the phosphopeptide (4), no peptide (5), the phosphopeptide derived from the corresponding region of JNK1&2 (6), and, the phosphopeptide derived from the corresponding region of ERK1&2 (7). After washing, membranes were incubated with goat F(ab')<sub>2</sub> anti-rabbit IgG HRP conjugate and bands were detected using the Pierce SuperSignal™ method. The data show that only the phosphopeptide corresponding to p38 [pTpY180/182] blocks the antibody signal, thereby demonstrating the specificity of the antibody.

