

## Monoclonal Antibody to EGF - Supernatant

<b>Alternate names:</b>	Beta urogastrone antibody, Epidermal Growth Factor antibody, Pro epidermal growth factor antibody, URG antibody, Urogastrone antibody
<b>Catalog No.:</b>	BM2207
<b>Quantity:</b>	1 ml
<b>Concentration:</b>	20 µg/ml
<b>Host / Isotype:</b>	Mouse / IgG1
<b>Clone:</b>	E5
<b>Format:</b>	<b>State:</b> Liquid supernatant <b>Buffer System:</b> 1% BSA and 20 mM Sodium Azide
<b>Applications:</b>	ELISA (10 ng EGF is detectable) and spot blots (1 ng is detectable). Use in formalin fixed frozen sections (1:20) and paraffin sections of salivary glands. See Protocol for more details. Other applications not tested. Optimal dilutions are dependent on conditions and should be determined by the user.
<b>Specificity:</b>	Reacts with EGF. In immunohistochemistry positive staining is observed in salivary glands. <b>Species:</b> Mouse. Does not work in Rat. Other species not tested.
<b>Storage:</b>	Store the antibody undiluted at 2-8°C for one month or (in aliquots) at -20°C for longer. Avoid repeated freezing and thawing. Shelf life: one year from despatch.
<b>General Readings:</b>	1. H.J. Beerstecher et al, 1988, J. of Histochemistry and Cytochemistry: vol. 36, 1153-1160.
<b>Protocols:</b>	Indirect Immunoperoxidase Staining On Frozen Sections : 1. 4 to 6 micron thick sections should be used. 2. Sections are thawed, 1-2 hours at room temperature. 3. Tissue is fixed in acetone, 10 minutes. 4. Wash with PBS, 2 x 3 minutes. 5. Incubate with monoclonal antibody (diluted in PBS), 1-2 hours at room temperature. 6. Wash with PBS, 3 x 3 minutes. 7. Incubate with peroxidase labeled second antibody, 30-60 minutes at room temperature. 8. Wash with PBS, 3 x 3 minutes. 9. Stain with diaminobenzidin (DAB) solution 10 minutes at room temperature. 10. Wash with running tap water, 3 minutes. 11. Counterstain with Mayer's hematoxylin, 2 minutes. 12. Wash with running tap water, 5 minutes. 13. Dehydrate with increasing solution of ethanol; 50%, 70%, 96%, absolute, 3 minutes

each.

14. Clear with xylol, 3 x 3 minutes.

15. Mount with mounting medium (e.g. malinol).

Indirect Immunoperoxidase Staining On Formalin-Fixed And Paraffin Embedded Tissues:

1. 4 micron thick sections should be used.

2. Dewax in Xylol, 3 x 3 minutes.

3. Rehydrate in decreasing grades of ethanol: absolute, 96%, 70%, 50%, 3 minutes each.

4. Block endogenous peroxidase activity with freshly made 0.3% H<sub>2</sub>O<sub>2</sub> in methanol, 20 minutes.

5. Wash with PBS, 3 x 3 minutes.

Only if trypsinisation is required

5a. Incubate sections with 0.1% Trypsin in 0.1% CaCl<sub>2</sub> pH 7.6 for 10 minutes at room temperature.

5b. Wash with PBS, 3 x 3 minutes.

6. Cover the sections with 20% normal rabbit serum in PBS or normal human serum and incubate overnight in a humidity chamber at room temperature to reduce non specific background staining.

7. Decant 20% normal rabbit serum.

8. Incubate with monoclonal antibody (diluted in PBS), 1-2 hours at room temperature.

9. Wash with PBS, 3 x 3 minutes.

10. Incubate with peroxidase labeled second antibody, 30-60 minutes at room temperature.

11. Wash with PBS, 3 x 3 minutes.

12. Stain with diaminobensidin (DAB) solution, 10 minutes at room temperature. A stock solution of 0.5% DAB in 0.5 M Tris/HCl (pH 7.4) can be made and stored frozen in the dark. Before use a quantity needed for staining can be thawed and diluted 10x with water. The diluted DAB solution should be filtrated. Just before use H<sub>2</sub>O<sub>2</sub> must be added to a final concentration of 0.01%.

13. Wash with running tap water, 3 minutes.

14. Counterstain with Mayer's hematoxylin, 2 minutes.

15. Wash with running tap water, 2 minutes.

16. Dehydrate with increasing solutions of ethanol: 50%, 70%, 96%, absolute, 3 minutes each.

17. Clear with xylol, 3 x 3 minutes.

18. Mount with mounting medium (e.g. malinol).