

**AM00301AF-N****Monoclonal Antibody to CDKN2C / p18INK4c - Azide Free**

<b>Alternate names:</b>	CDKN6, Cyclin-dependent kinase 4 inhibitor C, Cyclin-dependent kinase 6 inhibitor, p18-INK4c, p18-INK6, p18INK6
<b>Quantity:</b>	0.1 mg
<b>Concentration:</b>	1.0 mg/ml
<b>Background:</b>	The INK4 family of proteins consists of four members that block progression from the G1-to-S phase of the cell cycle by inhibiting the activity of Cdk4 and Cdk6. The p18INK4c cyclin-dependent kinase inhibitor is an important regulator of cellular differentiation and cell cycle progression, and it also acts as a potent tumor suppressor. p18INK4c is regulated by the transcription factors E2F1 and SP1 in response to environmental and intracellular signals such as cytokines, oncogenic overload, or cellular senescence.
<b>Uniprot ID:</b>	<a href="#">P42773</a>
<b>NCBI:</b>	<a href="#">NP_001253</a>
<b>GeneID:</b>	<a href="#">1031</a>
<b>Host / Isotype:</b>	Mouse / IgG2a
<b>Clone:</b>	DCS-118
<b>Immunogen:</b>	Bacterially produced His-tagged p18 proteins.
<b>Format:</b>	<b>State:</b> Liquid purified IgG fraction. <b>Purification:</b> Protein-A Sepharose Chromatography. <b>Buffer System:</b> PBS, pH 7.2 containing 50% Glycerol without preservatives.
<b>Applications:</b>	<b>Western Blot:</b> 1 µg/mL <i>Positive Control:</i> Saos-2 Cells. <b>Immunoprecipitation:</b> 3 µg/200-300 µL of cell extract. <i>Positive Control:</i> Saos-2 Cells. <b>Immunohistochemistry:</b> 1-5 µg/mL Heat treatment is necessary for Paraffin Embedded Sections. Microwave oven: 2 times for 10 minutes each in citrate buffer (pH 6.5). <i>Positive Control:</i> Tonsil Tissue. Detailed procedure is provided in <b>Protocols</b> . Other applications not tested. Optimal dilutions are dependent on conditions and should be determined by the user.
<b>Specificity:</b>	This antibody reacts with Human p18INK4c.
<b>Species Reactivity:</b>	<b>Tested:</b> Human.
<b>Add. Information:</b>	This product was originally produced by MBL International.
<b>Storage:</b>	Store the antibody undiluted at -20°C. Shelf life: one year from despatch.

**General Readings:**

1. Thullberg, M., et al. Hybridoma 19, 63-72 (2000)  
Clone DCS-118 is used in this reference.

**Protocols:****SDS-PAGE & Western Blotting**

- 1) Wash the cells 3 times with PBS and suspend with 10 volume of cold Lysis buffer (50 mM Tris-HCl, pH 7.2, 250 mM NaCl, 0.1% NP-40, 2 mM EDTA, 10% glycerol) containing appropriate protease inhibitors. Incubate it at 4°C with rotating for 30 minutes, then sonicate briefly (up to 10 seconds).
- 2) Centrifuge the tube at 12,000 x g for 10 minutes at 4°C and transfer the supernatant to another tube. Measure the protein concentration of the supernatant and add the Lysis buffer to make 8 mg/mL solution.
- 3) Mix the sample with equal volume of Laemmli's sample buffer.
- 4) Boil the samples for 2 minutes and centrifuge. Load 10 µL of the sample per lane in a 1 mm thick SDS-polyacrylamide gel for electrophoresis.
- 5) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm<sup>2</sup> for 1 hour in a semi-dry transfer system. (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% MeOH). See the manufacture's manual for the transfer procedure.
- 6) To reduce nonspecific binding, soak the membrane in 10% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature, or overnight at 4°C.
- 7) Incubate the membrane with the anti-p18INK4c (DCS-118) monoclonal antibody (1 µg/mL) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature.
- 8) Wash the membrane with PBS (5 minutes x 6 times).
- 9) Incubate the membrane with the 1:10000 HRP-conjugated anti-mouse IgG diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature.
- 10) Wash the membrane with PBS (5 minutes x 6 times).
- 11) Wipe excess buffer from the membrane, then incubate it with appropriate chemiluminescence reagents for 1 minute. Remove extra reagent from the membrane by dabbing with a paper towel, and seal it in plastic wrap.
- 12) Expose to an X-ray film in a dark room for 5 minutes. Develop the film as usual. The conditions for exposure and development may vary.

Positive Control for Western blotting: Saos-2 cells.

**Immunoprecipitation**

- 1) Wash the cells 3 times with PBS and suspend with 10 volume of cold Lysis buffer (50 mM Tris-HCl, pH 7.2, 250 mM NaCl, 0.1% NP-40, 2 mM EDTA, 10% glycerol) containing appropriate protease inhibitors. Incubate it at 4°C with rotating for 30 minutes, then sonicate briefly (up to 10 seconds).
- 2) Centrifuge the tube at 12,000 x g for 10 minutes at 4°C and transfer the supernatant to another tube.
- 3) Add 3 µg of the anti-p18INK4c (DCS-118) monoclonal antibody into 250 µL of the supernatant. Mix well and incubate with gentle agitation for 30-120 minutes at 4°C. Add 20 µL of 50% Protein A-agarose beads resuspended in the Lysis buffer. Mix well and incubate with gentle agitation for 60 minutes at 4°C.
- 4) Wash the beads 3-5 times with ice-cold Lysis buffer (centrifuge the tube at 2,500 x g for 10 seconds).
- 5) Resuspend the beads in 20 µL of Laemmli's sample buffer, boil for 3-5 minutes, and centrifuge for 5 minutes. Use 10 µL/lane for the SDS-PAGE analysis.

(See **SDS-PAGE & Western blotting.**)

Positive Control for immunoprecipitation: Saos-2 cells.

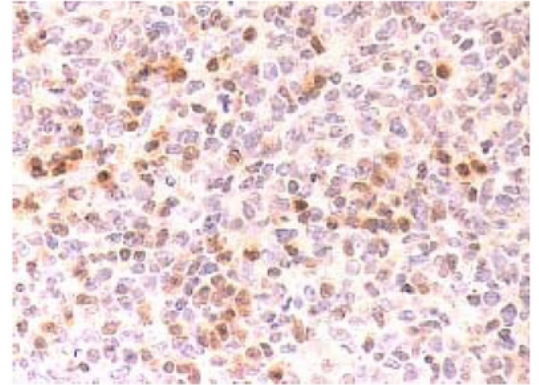
**Immunohistochemical Staining for Paraffin-Embedded Sections: SAB method**

- 1) Deparaffinize the sections with Xylene 3 times for 3-5 minutes each.
- 2) Wash the slides with Ethanol 3 times for 3-5 minutes each.
- 3) Wash the slides with PBS 3 times for 3-5 minutes each.
- 4) Heat treatment  
Heat treatment by microwave oven: Place the slides put on staining basket in 500 mL beaker with 500 mL citrate buffer (pH 6.5). Cover the beaker with plastic wrap, then process the slides 2 times for 10 minutes each at 500 W with microwave oven. Let the slides cool down in the beaker at room temperature for about 40 minutes.
- 5) Remove the slides from the citrate buffer and cover each section with 3% H<sub>2</sub>O<sub>2</sub> for 10 minutes at room temperature to block endogenous peroxidase activity. Wash 3 times in PBS for 5 minutes each.
- 6) Remove the slides from PBS, wipe gently around each section and cover tissues with Protein Blocking Agent for 5 minutes to block non-specific antibody staining. Do not wash.
- 7) Tip off the blocking buffer, wipe gently around each section and cover tissues with the anti-p18INK4c (DCS-118) monoclonal antibody diluted with PBS containing 1% BSA (1-5 µg/mL).
- 8) Incubate the sections for 1 hour at room temperature.
- 9) Wash the slides 3 times in PBS for 5 minutes each.
- 10) Wipe gently around each section and cover tissues with Polyvalent Biotinylated Antibody. Incubate for 10 minutes at room temperature. Wash as in step 9.
- 11) Wipe gently around each section and cover tissues with Streptavidin-Peroxidase. Incubate for 10 minutes at room temperature. Wash as in step 9.
- 12) Visualize by reacting for 10-20 minutes with substrate solution containing 7.5 mg DAB, 40 µL of 30% H<sub>2</sub>O<sub>2</sub> in 150 mL PBS. \*DAB is a suspected carcinogen and must be handled with care. Always wear gloves.
- 13) Wash the slides in water for 5 minutes.
- 14) Counter stain in hematoxylin for 1 minute, wash the slides 3 times in water for 5 minutes each, and then immerse the slides in PBS for 5 minutes. Dehydrate by immersing in Ethanol 3 times for 3 minutes each, followed by immersing in Xylene 3 times for 3 minutes each.
- 15) Now ready for mounting.

Positive Control for Immunohistochemistry: Tonsil Tissue.

**Pictures:**

**Figure 3.** AM00301AF-N p18INK4c antibody Staining of Paraffin Embedded Human Tonsil Section (germinal center).



**Figure 1.** Western blot analysis of p18INK4c expression in Saos-2 cells (lane 1), C2C12 cells (Lane 2) and Rat-1 cells (Lane 3) using p18INK4c antibody (AM00301AF-N). **Figure 2.** Immunoprecipitation of p18INK4c from Saos-2 cells with normal Mouse IgG (Lane 1) or AM00301AF-N (Lane 2). After immunoprecipitated with the antibody, immunocomplex was resolved on SDS-PAGE and immunoblotted with p18INK4c antibody (AM00301AF-N).

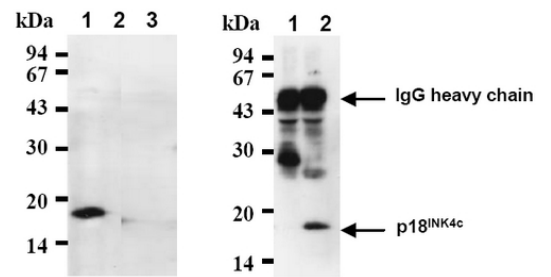


Figure 1.

Figure 2.